



## SCREENING OF DYE DEGRADING BACTERIA FROM TEXTILE EFFLUENTS

T.Vinodhkumar<sup>1</sup>,  
N.Thiripurasundari\*<sup>1</sup>,  
G.Ramanathan<sup>2</sup>, G. Karthik<sup>3</sup>

<sup>1,3</sup> Department Of Microbiology,  
Avscollege Of Arts And Science,  
Salem, Tamil Nadu, India

<sup>2</sup>department Of Microbiology,  
V.H.N.S.N. College,  
Virudhunagar-626 001, India

### ABSTRACT

The textile industry in India is one of the oldest industries. It provides direct employment to nearly thirty million people. Wastewater from textile industries are a complex mixture of many polluting substances such as organochlorine based pesticides, heavy metals, pigments and dyes. This study deals about the biodegradation of Reactive yellow 81, which is a commercially important dye, with a wide range of applications across the textile and leather industries. Decolourisation was assessed as color disappearance during bacterial growth. Textile effluent samples were collected from different dyeing units near Salem. Seventeen isolates were isolated. Using plate assay method the decolourisation activity of bacteria was detected. Among the seventeen isolates seven organisms showed the decolourisation activity and they are *Bacillus* sp 1, *Staphylococcus*, *Kurthia*, *Legionella*, *Pseudomonas*, *Sulfidobacillus*, *Bacillus* Sp.

**Key words** Textile Industry, Dye, Degradation, Effluent

## INTRODUCTION

Environmental pollution has been defined in various ways. It is considered as the release of unwanted substances to the environment by man in quantities that damage either the health or the resources itself. Water pollution involves the release of small amounts of substances directly through point sources or indirectly through nonpoint sources. Industrial effluents from various industries like textile, dye stuffs, paper and pulp, distillery, Olive oil mill and metal industries are the major contributor to water pollution.

A dye can generally be described as a colored substance that has an affinity to the substrate to which it is being applied. The dye is generally applied in an aqueous solution, and may require a mordant to improve the fastness of the dye on the fiber.

Dyes are widely used in the textile, rubber product, paper, printing, color photography, pharmaceuticals, cosmetics and many other industries. Waste water from textile industries creates a great pollution problem due to the dye content. Industrialization is vital to a nation's economy because it serve as a vehicle for development. However, there are associated problems resulting from the introduction of industrial waste products into the environment. Many of these products are problematic because of persistence (low biodegradability) and / or toxicity.

India's dye industry produces every type of dyes and pigments. Production of dyestuff and pigments in India is close to 80,000 tones. India is the second largest exporter of dye stuffs after china. The textile industry accounts for the largest consumption of dye stuffs (nearly 80%).

The Indian textile industries now predominantly use synthetic organic dyes like direct dyes, processing dyes, reactive dyes etc. The large variety of dyes and chemicals used in an attempt to make more attractive popular shades of fabrics for a competitive market render them very complex. During the last decade, environmental issues associated with dye stuff production and application have grown significantly and are indisputably among the major driving forces affecting the textile dye industry today.

Management of water pollution is at present one of the major challenges for environmentalists. More than 10,000 different textile dyes with an estimated annual production of  $7 \times 10^5$  metric tones are commercially available worldwide (Mc Mullan et al., 2001). 2% of these dyes are directly discharged as aqueous effluents and 10% are subsequently lost during textile coloration process (Pearce et al., 2003). Color is one of the most obvious indicators of water pollution, and discharge of highly colored synthetic dye effluents can be damaging to the receiving water bodies (Nigam et al., 1996).

### Reactive Dyes

Reactive dye is a class of highly colored organic substances, primarily utilized for tinting textiles that attach themselves to their substrates by a chemical reaction that forms a covalent bond between the molecule of dye and that of the fiber. In this reactive dye cold brand dyes, hot brand, vinylsulphone based, highly exhausted and printing dyes are present. "Cold" reactive dyes such as procion Mx, Cibacron F, and Drimarenek, are very easy to use because they can be applied at room temperature. Reactive dyes are by far the best choice for dyeing cotton and other cellulose fibers at home or in the art studio.

### Impact of Textile Effluents in Environment

The textile industries produce effluents that contain several types of chemicals such as dispersants, leveling agents, acids, alkalis, carriers and various dyes (Cooper, 1995). The release of colored compounds into water bodies is undesirable not only because of their impact on photosynthesis of aquatic plants but also due to the carcinogenic nature of many of these dyes and their breakdown products (Weisburger, 2002). This alters the  $P^H$ , increases the biological oxygen demand (BOD) and chemical oxygen demand (COD) and gives the rivers intense colorations (Ajayi and Osibanjo, 1980). The use of these water resources is limited and the ecosystem is affected.

### Treatment of Dye effluent

The textile industry in India is one of the oldest industries. It provides direct employment to nearly thirty million people. Wastewater from textile industries are a complex mixture of many polluting substances such as organochlorine based pesticides, heavy metals, pigments and dyes.

The wastewater characteristics from a dye house are highly variable from day to day, depending on the type of dye, the type of fabric and the concentration of the agents added. Treatment of such wastewaters is therefore, essential but difficult. The removal of dyes from the textile waste effluent has been carried out by physical, chemical and biological methods such as flocculation, membrane filtration, electrochemical techniques, ozonation, coagulation, adsorption and fungal discoloration. (Fu et al., 2001). In recent years a number of studies have focussed on some microorganisms which are able to biodegrade and biosorb dyes in waste waters.

### Role of Microbes:

A wide variety of microorganisms, capable of decolorizing a wide range of dyes include some bacteria fungi and algae (Fu et al., 2001). The use of microorganisms for the removal of synthetic dyes from industrial effluents offers considerable advantages. The process is relatively inexpensive, it is simple method and the running costs are low. The end products of complete mineralization are not toxic (Zheng et al, Forgacs et al and Park et al., 2004).

Biodegradation processes may be anaerobic, aerobic or involve a combination of the two (Forgacs et al., 2004). However, it has been observed a number of cases that the efficiency of aerobic treatment was inferior to that of anaerobic decolorization process. Although anaerobic reduction of azo dyes is generally more satisfactory than aerobic degradation, the intermediate products (carcinogenic aromatic amines) have to be degraded by an aerobic process.

Bacteria involved in biodegradation of dyes are *Bacillus firmus*, *Bacillus laterosporus*, *Legionella*, *Chryseobacterium*, *Flavobacterium*, *Pseudomonas*, *Enterobacter* sp, *Serratia*, sp *Yersinia* sp, *Erwinia* sp, and *Bacillus subtilis*. Some of the fungus such as *Bjerkandera* sp, *Schizophyllum commune*, *Penicillium oxalicum*, *Rhizopus arrhizus*, *Phanerochaete chrysosporium*, *Fusarium moniliforme*, *A.terreus*, *A.niger*, *Mucor racemosus*, *Cladosporium*, *cladosporioides*, *Trichoderma viride*, *A.ochraceus*, Thermotolerant yeast, *Kluyveromyces marxianus* are involved in dye degradation. Algae such as

Nostoc muscorm, Scenedesmus bijugatus, Cyanobacteria, and diatom Nitzschia perminuta are also take part in dye degradation. Biological degradation of dye effluent is evaluated as a good treatment method.

This study is aimed to asses the potential of bacteria isolated from textile effluent in Salem district to degrade the dye reactive yellow81 (yellow HE4G) is chosen for this degradation study, as it is one of the frequently used dye in dyeing industry in Salem.

## **MATERIALS AND METHODS**

### **Collection of samples**

10 Samples from dye house effluent were collected from a dyeing unit in Salem, Tamil Nadu, India. They were stored in refrigerator at 4<sup>0</sup>C and used without any pretreatment.

### **Dye**

The dye used for the degradation study is Reactive yellow 81 (yellow HE4G). It is a bi functional monochlorotriazine dye of the azo class. The molecular formula of this dye is C<sub>52</sub> H<sub>34</sub> Cl<sub>2</sub> N<sub>180</sub> 20S<sub>6</sub>. 6Na.

### **Structure of Reactive yellow 81 (HE4G)**

### **Analysis of Physiochemical Parameters:-**

Physiochemical parameters of the collected textile dye effluent such as P<sup>H</sup>, color and odour were measured using standard methods.

### **Isolation of bacteria from Textile Effluent**

The textile effluent is serially diluted and spread plated in nutrient agar. An each type of isolated colonies were subculture for their enrichment and stored in agar slant.

### **Identification of the Isolates**

A pure colony of the unknown isolates identified presumptively on the basis of the following tests Gram staining Method, Spore staining, Motility Test, Catalase Test, Oxidase Test, Nitrate Test, Gelatin hydrolysis.

**SCREENING OF DYE DECOLOURISING MICROORGANISM (HUSSEINY Sh.M.)****Inoculum Preparation**

The 24 hours old culture of the bacteria was used to investigate their ability to decolorize the reactive yellow 81.

**Plate Assay Method**

Plate assay was performed for the detection of decolorizing activity of bacteria. The nutrient agar and reactive yellow 81 dyes were autoclaved at 121°C for 15 minutes. The isolated organisms were plated on nutrient agar, plates containing reactive yellow 81 (500mg /l). The plates were wrapped with parafilm and were incubated at 37°C for 7 days. The plates were observed for clearance of the surrounding the colonies.

**Medium used for Decolourization Test**

The experiments were carried out in 100 ml flasks containing 50 ml of the tested reactive yellow 81 dye solution in the concentration of 500 mg/L, trace of yeast extract and sucrose. The P<sup>H</sup> was adjusted to 7 ± 0.2 using diluted Na<sub>2</sub>CO<sub>3</sub> and HCL solutions.

Then two sets of dye solution were prepared for each isolate inoculated with 1ml of inoculum of each bacterial isolate. The flasks were incubated at 37°C for 12 days one set of the dye solution maintained in stationary condition and another one in shaking incubated to identify the effectiveness of dye degradation.

At 24 hours interval the samples were analyzed for degradation. 5 ml of the test dye solution was centrifuged at 5000 rpm for 15 minutes and the optical density of the supernatant was measured by spectrophotometry at  $\lambda$  max (570 nm) of the reactive yellow 81.

**Analytical Methods:**

Absorbance of the supernatant withdrawn at 24 hrs intervals were measured at the maximum absorption wavelength ( $\lambda$  max - 570 nm) for reactive yellow 81 in the visible region on Spectrophotometer. The percentage of decolourisation was calculated.

**RESULTS****Analysis of Physiochemical Parameters**

10 different textile dye effluent samples were collected from a dyeing unit in Salem. P<sup>H</sup> of the all textile effluent was 7.5. As the dye effluent was collected from different sites, it was having different color (Green, Orange, Pink, Red, Blue) based on the dye which was used for dyeing in the dyeing unit. There was no odor in the effluent samples.

### Isolation of Bacteria from Textile Effluent

Well isolated colonies were obtained by spread plate technique. Mixed populations of organisms were separated by this technique and discrete colonies were isolated. A total of 7 different isolates were obtained and these isolates were used for decolorization studies.

### Plate assay method screening of Dye Decolorizing Microorganism

Decolorizing ability of the 7 isolates were identified by using the Reactive Yellow 81 dye which is a commercially important dye, with a wide range of applications across the textile and leather industries.

Dye solution inoculated with the isolates was maintained in both stationary and agitated conditions separately.

Optical density of the dye solution inoculated with the isolates for both stationary and agitated conditions was taken separately and are given in table 4 and 5 respectively.

The percentage of dye decolourization of the isolates in both stationary and agitated condition was calculated with a standard formula and are given in the table 6.

S.No	Isolates	% of Decolourization	
		Under Stationary Condition	Under Agitated Condition
1	Pseudomonas sp	37.5 %	25 %
2	Staphylococcus sp	25 %	12.5 %
3	Kurthia sp	12.5 %	12.5 %
4	Legionella sp	37.5 %	37.5 %
5	Sulfidobacillus sp	37.5 %	25 %
6	Bacillus sp1	50 %	37.5 %
7	Bacillus sp2	12.5 %	25 %

**Table 1 :Decolourization of Reactive Yellow 81 by the Isolates**

## DISCUSSION

A total of 17 isolates were obtained from textile effluent samples. Among which five isolates belonging to the genera *Bacillus* (from sample 1,2,5,6,3) and four isolates are *Staphylococcus* (from sample 1,3,5,6). Three isolates belonging to the genus *Sulfidobacillus* (from sample 7,8) and one genera of *Kurthia* (from sample 5) and *Legionella* (from sample 7,8) three genera of *Pseudomonas*(from 7.8.5) were isolated from the textile effluents.

From the dye contaminated soil, *Bacillus* was isolated dominantly by Leena et al., (2008). The other bacteria such as *Pseudomonas*, *Arthrobacter*, *Alcaligenes* were also isolated.

Thus among the all isolates, *Bacillus* is the Predominant one, which is largely isolated from the textile effluent. Other than this *Bacillus* is also an effective biodegrader of various dyes such as pigmented red 208 Saraswathi et al., (2009). Reactive Black-B Leena et al., (2008), Acidblue 113 Gurulakshmi et al., (2008), Reactive red (RR) Barakat et al., (2009), Cibacronblack PSG, cibacron red P4B Ola et al., (2010).

In this study, the decolourisation ability of the isolates against reactive yellow 81 was demonstrated. The optical density was measured at 570nm with two days interval for 12 days. On the Second day *Pseudomonas* showed 25% decolourisation, *Bacillus* 1 showed 12.5%. Whereas other isolates did not show any decolourisation. *Legionella* showed 25% decolourisation during the fourth day and the other isolates such as *Pseudomonas* (25%) *Bacillus* 2, *Sulfidobacillus*, *Bacillus* 1 showed 12.5% decolourisation. *Staphylococcus* showed 12.5% and *Bacillus* 1 has 37.5% on sixth day. Followed by further incubation on eight day except *pseudomonas* (37.5 %) *sulfidobacillus* (25%) other isolates does not show any decolourisation on Ten day 50 % decolourisation was showed by *Bacillus* 1, *Kurthia* shows 12.5 % only on this day. Finally 25 % decolourisation was observed in *Staphylococcus*, *Legionella* and *sulfidobacillus* has 37.5 %, *Bacillus* 2 remains the same as it was in the sixth day. These conditions were observed under stationary condition.

*Bacillus* 2 has 12.5% decolourisation from second, fourth, sixth, eighth, day and it has 25% decolourisation on tenth and 12th day.

Compare to stationary and agitated condition the isolates exhibited effective dye decolorizing activity only when incubated under the stationary conditions. Whereas negligible decolourisation was noticed under the agitating conditions. the further incubation of these isolates on reactive yellow 81 did not improve decolorization. Anaerobic or stationary conditions were necessary for bacterial decolourisation though the cell growth was poorer than that under aerobic conditions. (Gurulakshmi et al.,2008). Among all the isolates *Bacillus* 1 is the first effective degrader of reactive yellow 81, secondly by *Legionella* next to them are *Pseudomonas*, *Sulfidobacillus*, *Bacillus* 2 and *Staphylococcus*, *Kurthia* sp. has the lowest decolourisation ability(Gurulakshmi, et al.,2008) stated that there was no increase in decolourisation of *Kurthia* whenever the Inoculum size is increased.

## BIBLIOGRAPHY

1. Abbash Attia, J., Salih Kadhim H, Falah Hussein H. Photocatalytic degradation of textile dyeing waste water using titanium dioxide and Zincoxide. *E -J of chemistry* 2008; 5, 2 : 219-223.
2. Abou-Okeil, A. Modified saw dust for dye removal, 2<sup>nd</sup> international conference of textile research division, NRC, cairo, Egypt 2005; April, 11-13.
3. Ajayi SO, Osibanjo O. The state of environment in Nig. Pollution studies of textile industries in Nig. Monogra 1980;1: 76-86.
4. Ajibola VO, Oniye SJ, Odeh CE, Olugbodi T and Umeh UG. Biodegradation of Indigo containing Textile Effluent using some strains of Bacteria *J of Applied Sciences* 2006; 5(5): 853-855.
5. Azeen khalid, Muhammad Arshad and David Crowley. Bioaugmentation of azo dyes. *Hdb Env chem.* 2010; 9: 1-37.
6. Bafana A, Chakrabarti T, Devi SS. Azoreductase and dye detoxification activities of *Bacillus velezensis* strain AB. *Appl Microbiol Biotechnol* 2008; 77(5):1139-44.
7. Barakat OS, Darwesh OM, Sedik MZ, Moawad H and Abd El-Rahim WM. Evidence of Biodegradation of reactive red textile azodye in anoxic / aerobic bioremediation system. 4<sup>th</sup> conference on Recent Technologies in Agriculture 2009.
8. Carliell, C.M., Barclay, S.J., Naidoo, N., Buckley, C.A., Mulholland D, A and senior. E 1994. Anaerobic decolorisation of reactive dyes in conventional sewage treatment processes. *Water SA* 20:341-344 .
9. Cheunbarn, T., Cheunbarn, S., and Khumjai, T 2008. Prospectus of bacterial granule for treatment of real textile industrial wastewater. *Int.J.Agr.Biol*,10:689-92.
10. Faraco, V., Pezzella, C., Giardina, P., Piscitelli, A., Vanhulle, S and Sannia, G 2009. Decolourization of textile dyes by the white rot fungi *phanerochaete chrysosporium* and *pleurotus ostreatus* *J chem Technol Biotechnol*, 84: 414-419.
11. Forgacs, E., Cserhati, T. and Oras, G 2004. Removal of Synthetic dyes from waste waters, *Environment International*, 30: 953-971.
12. Franciscon Elisangela, Zille Andrea, Dias Guimaro Fabio, Ragagnin de Menezes Cristiano, Durrant Lucia Regina, Covaco. Paulo Artur. 2009. Biodegradation of textile azo dyes by a facultative *staphylococcus arlettae* strain VN-II using a sequential microaerophilic / aerobic process. *International Biodeterioration & Biodegradation* 63: 280-288.
13. Fu, Y, and Tiraraghavan, Y 2001. Fungal decolourization of dye waste waters: a review, *Bioresource Technology*, 79: 251-262.
14. Ganesh Parshetti, Satish kalme, Ganesh Saratale, and Sanjay Govindwar 2006. Biodegradation of Malachite Green by *Kocuria rosea* MTCC 1532. *Acta chim. Slov* 53 :492-498.
15. Georgiou, D., Dinske, J., Metallinou, C and Aivasidis, A 2003. Decolorization of Azo – reactive dyes and cotton textile wastewater using anaerobic digestion and acetate consuming bacteria.
16. Griffiths, J. 1984 Developments in the light absorption Properties of dyes colour and Photochemical degradation reactions. In: *Developments in the chemistry and Technology of organic dyes* Griffith J (Eds). Oxford: Soc.chem Ind, :1-30.
17. Gurulakshmi, M., Sudarmani, D.N.P., Venba, R 2008. Biodegradation of Leather Acid dye by *Bacillus Subtilis*. *Advanced Biotech* : 12-18.
18. Hanan Hafez omar. 2008. Algal decolorization and degradation of monoazo and diazo Dyes. *Pak.J. Biol. Sci.*, 11: 1310-1316.
19. Husseiny, Sh.M 2008. Biodegradation of the reactive and direct dyes using egyptian isolate *J Appl.sci.Res.*, 4(6): 599-606.
20. Idil Arslan Alaton, Guclu Insel, Gulen Eremektar, Fatos Germirli Babuna, Derin Orhon. 2006. Effect of textile auxiliaries on the biodegradation of dye house effluent in activated sludge chemosphere 62: 1549-1557.



21. Kalyanee Jirasripongpun, Rujikan Nasanit, Jongjira Niruntasook, and Boonsiri Chotikasatian . 2007. Decolorization & degradation of C.I. reactive red 195 by *Enterobacter* sp. *Thammasat Int.J.SC.Tech.*, 12, No.4.
22. Khadijah, O., Lee, K.K., and Mohd faiz, F., Abdullah. 2009. Isolation, Screening and development of local bacterial consortia with azodyes decolourising capability. *Malaysian J of Microbiology*, 5(1): 25-32.
23. Kodam, K.M. and Gawai, K.R 2006. Decolorisation of reactive red 11 and 152 azo dyes under aerobic conditions. *Indian J of Biotechnol*, 5:422-424.
24. Kulkarni, S.V., Blackwell, C.D., Blackard A.L., Stackhouse, C.W and Alexander, M.W 1985. Textile dyes & dyeing equipment classification, properties & environmental aspects.
25. Ladhari, N., Hayet, G. and Harrabi.L 2001. study of the Biodegradability of Sodium Salt from the condensation Product of naphthalene sulphonic acids and formaldehyde file:///H:/New% 20 folder/Textile%20 Research% 20% 20 study % 2 ....
26. Leena, R and Selva Raj, D 2008. Biodecolourization of textile effluent containing Reactive Black-B by effluent – adapted and non-adapted bacteria. *African J of Biotechnology*, 7(18): 3309-3313.
27. Li, J and Bishop, P.L 2002. Insitu identification of azo dye inhibition effects on nitrifying biofilms using microelectrodes. *Water science and Technology* 46 No 1-2: 207-214.
28. Maria Jonstrup and Bo Mattiasson. 2008. Development of Process technology for treatment of textile wastewaters. U21 PostGraduate Research conference proceedings1.
30. Mathur, N., Bhatnagar P., Bakre,P 2005 Assessing mutagenicity of textile Dyes from pali (Rajasthan) using AMES Bioassay. *Applied Ecology and Environmental Research* 4(1): 111-118.
31. McMullan, G., Mehan, C., Conneely,A., Kirby, N., Robinson,T., Nigam,P., Banat, I.M., Marchant,R., Smyth W.F 2001. Microbial decolourization and degradation of textile dyes. *Appl. Microbiol Biotechnol.* 56: 81-87.
32. Muthazhilan, R., Yogananth, N., Vidhya,S. and Jayalakshmi, S 2008. Dye Degrading Mycoflora from Industrial Effluents. *Res.J.Microbiol.*,3(3): 204-208.
33. Nigam,P., Banat, I.M., Singh, D., Marchant, R 1996. Microbial Process for the decolourization of textile effluent containing azo, diazo and reactive dyes. *Process Biochem.* 31: 435-442.
34. Olukanni, O.D., osuntoki, A.A., and Gbente, G.O 2006. Textile effluent biodegradation potentials of textile effluent – adapted and non-adapted bacteria. *Afr. J.Biotechnol*, 5(20): 1980-1984.
35. Olukani,O.D.,Osuntoki, B.V. and upadhyay, R.S 2010. *Pseudomonas fluorescens* can be used for bioremediation of textile effluent Direct Orange – 102. *Tropical Ecology* 51(23): 397-403.
36. Park, C.h., Lee, M., Lee,B., Kim, S.W., Chase, H.A., Lee,J., and Kim,S 2006. Biodegradation and biosorption for decolourization of synthetic dyes by *funaliatrogii*, *Journal of Biochemical Engineering*, In Press,: 1-17.
37. Pearce,C.I., Lioyd,J.R., Guthrie, J.T 2003. The removal of colour from textile wastewater using whole bacterial cells: a review. *Dyes pigments* 58: 179-196.
38. Pigment-wikipedia,the free encyclopedia. file:///G:/pigment.htm.

39. Puvaneswari, N., Muthukrishnan, J., and Gunasekaran, P 2006 Toxicity assessment and microbial degradation of azodyes Indian J Exp Biol, 44: 618-626.
40. Ramesh babu, B., Parande, A.K., Raghu, S. and Prem Kumar, T 2007. Cotton Textile processing ; Waste Generation and Effluent Treatment. The J of cotton science 11 : 141 – 153.
41. Reactive dyes. file : [/// D: / DOWNLOAD% 20 FILES / reactive – dyes. htm](#).
42. Saleh El-Kadri, Ousama Dabbit and Hassan Kakhia.2002. Treatment of wastewater containing dyes used in the Syrian textile industry. J chem Technol Biotechnol 77: 437-443.
43. Sanroman, M.A., Deive, F.J., Dominguez, A., Barrio, T., Longo, M.A 2009. Dye decolourization by nearly isolated thermophilic microorganisms. [http : // bioprocesos. Uvigo.es/](http://bioprocesos.Uvigo.es/).
44. Saraswathi, K. and Balakumar, S 2009. Biodecolourization of azodye (pigmented Red 208) using *Bacillus firmus* and *Bacillus laterosporus*. J.Biosci Tech, 1(1): 1-7.
45. Shailesh, B., Siddharth,J., Gondaliya, Pathak 2007. Kinetic of Azodyes decolourization by Enterobacteriaceae species in the Intact cell Assay system J.Appl.Sci. Environ, Manage. 11 (3) 45 – 52.
46. Suresh,K., Prakash, D., Rastogi.N and Jain, R.K 2007. *clostridium nitrophenolicum* sp nov., a novel anaerobic P-nitrophenol-degrading bacterium, isolated from a subsurface soil sample. Int J syst Evol Microbiol 57: 1886-1890.
47. Syed, M.A.,Sim, H.K.,Khalid, A and shukor, M.Y 2009. A simple method to screen for azo-dye degrading bacteria. J Environ. Biol, 30(1): 89-92.
48. Ten, L.N., Im, W.T., Kim, M.K., Lee, S.T 2004. A plate assay for simultaneous screening of polysaccharide and protein degrading microorganisms. Letters in Applied Microbiology.40: 92-98.
49. Veeranagouda, Y., Neelakanteshwar patil, K., Karegoudar, T.B 2004. A method for screening of bacteria capable of degrading dimethylformamide. Current science 87 No.12.
50. Weisburger, J.H 2002. Comments on the history and importance of aromatic and heterocyclic amines in public health. Mutat. Res. 506/507: 9-20.
51. Yongjie Miao.2005. Biological remediation of Dyes in textile effluent: a review on current treatment technologies.
52. Zheng, Z., Levin, R.E., Pinkhm, J.L and Shetty,K 1999. Decolourization of Polymeric dyes by a novel *Penicillium* Isolate, Process Biochemistry, 34: 31-37.